

## Chapter 3

# The physiological basis of ozone injury assessment attributes in Sierran conifers

Nancy E. Grulke

*USDA Forest Service, Pacific Southwest Research Station, 4955 Canyon Crest Dr.,  
Riverside, CA 92507-6099, USA  
E-mail: ngrulke@fs.fed.us*

### Abstract

The generalized effects of ozone ( $O_3$ ) exposure on plants are well known, but translating responses known under controlled conditions to effective tools for assessments under field conditions can be complex. There are two methods generally used for assessing yellow pine (ponderosa and Jeffrey) response to oxidant exposure in the Sierra Nevada: the Ozone Injury Index (OII) and the Forest Pest Management (FPM) assessment. For both methods, chlorotic mottle and needle retention form the basis of the assessment. The response of these two key attributes to cumulative  $O_3$  exposure and uptake, nitrogen deposition (either associated with  $O_3$  deposition as a correlative environmental factor or as an experimental manipulation), and drought stress are described using field examples from both ponderosa and Jeffrey pine. Specific suggestions are offered for future, repeated field assessments of  $O_3$  injury for consistency, and to help interpret historical data sets.

### 1. Introduction

The generalized effects of ozone ( $O_3$ ) exposure on plants are well known, and have been reviewed from several viewpoints over the last decade (De Kok and Tausz, 2001; Heath and Taylor, 1997; Pell et al., 1997; Schraudner et al., 1997; Matyssek et al., 1995; Bytnerowicz and Grulke, 1992). In general, oxidant exposure results in lower photosynthetic capacity, higher respiration, lower stomatal conductance, lower nutritional content of tissues, as well as elemental imbalances. Oxidant exposure alters within plant priorities for resources: less carbon is allocated to roots, and less foliar biomass is retained.

The specific effects of  $O_3$  exposure on conifers are well described (Darrall, 1989; Reich, 1987). Most of the research on  $O_3$  effects has been conducted on herbaceous species (i.e., crops). Several important differences between herbaceous and coniferous species affect the total foliar uptake of the pollutant, the

timing of the uptake, and the effect of multiple stressors. However, the underlying mechanisms of O<sub>3</sub> injury response in conifers and herbaceous species are assumed to be similar.

One of the most important differences between herbaceous and conifer species relevant to oxidant exposure is lower stomatal conductance. Stomatal conductance in conifers is roughly half that of herbaceous species, so that at the same O<sub>3</sub> exposure level, O<sub>3</sub> uptake in conifers is proportionately reduced. However, conifer foliage is generally longer-lived, and active over a greater portion of the year. A longer period of physiological activity increases the total O<sub>3</sub> uptake over the course of the year, and greater foliar longevity increases total O<sub>3</sub> exposure over the lifetime of the leaf. Conifers are physiologically active in early spring and late fall, during times of lower oxidant concentrations. These periods can contribute significantly to a net positive carbon balance over the course of the year. Patterson and Rundel (1995) reported that Jeffrey pine had significant stomatal opening (one third that of a summer day) in mid winter with 2 m of snow on the ground. At least pole-sized and larger trees can mitigate reductions in carbon acquisition due to oxidant exposure in the summer with carbon assimilation on favorable days in the winter.

Conifers have thicker cuticles relative to herbaceous species. Although cuticular uptake of O<sub>3</sub> is believed to be negligible (Kerstiens and Lenzian, 1989), other associated pollutants (e.g., HNO<sub>3</sub>) in the southern Sierra Nevada and in the Transverse Range may compromise cuticular integrity (Bytnerowicz et al., 1999) and contribute to the total stress experienced. Needle longevity may be an advantage from the point of view of nutrient storage important in reparation responses to pollutants. However, cuticular lesions may be more common in older needles and trans-cuticular loss of water is unlinked to plant carbon gain. The potential for "leaky" cuticles to contribute to whole canopy pollutant uptake in mature trees (not to be confused with pollutant deposition to foliar surfaces) has yet to be quantified under field conditions.

The longevity of the whole tree also holds advantages and disadvantages for response to pollutants. Greater longevity of conifers translates into greater environmental volume from which to scavenge resources, a greater proportion of resources carried over from year to year and the potential to buffer the individual from environmental stressors and allocate more resources to plant defense (antioxidants). Conversely, longevity can also work against the tree in the form of multiple, cumulative stressors. A series of drought years can reduce O<sub>3</sub> uptake, but can also reduce carbon and nutrient acquisition, reducing resource allocation to defenses (antioxidants, resins against insect infestation), rendering the tree more susceptible to O<sub>3</sub> injury in a subsequent year of greater precipitation, greater stomatal conductance, greater O<sub>3</sub> uptake, and injury. These differences in resource acquisition and allocation all contribute to altering carbon and nitrogen dynamics within the tree.

Nitrogen deposition modifies the effects of oxidant exposure. Nitrogen deposition, in wet or dry particulate form, ultimately increases site fertility. This can occur to excess (“N saturation”) as has been shown by Fenn et al. (1996). Increased nitrogen availability decreases carbon allocation to roots, further exacerbating the effects of oxidant exposure on roots. Increased nitrogen availability also exacerbates O<sub>3</sub> exposure effects on foliage turnover: fewer needle age classes are retained (Gower et al., 1993). Nitrogen amendments counteract the effect of oxidant exposure on photosynthesis by increasing nitrogen available for photosynthetic pigments and enzymes, increasing stomatal conductance, but deleteriously resulting in increased O<sub>3</sub> uptake. On the other hand, nitrogen amendments may mitigate the degree of foliar injury from oxidant pollution via higher nitrogen available for reparation of photosynthetic pigments. Nitrogen amendments also modify the antioxidant defense system in complex ways (Polle, 1998). In addition to these effects, estimated stomatal uptake of gaseous nitrogen oxides may be significant (Bytnerowicz et al., 1996) and phytotoxic (Tausz, 2001).

Over much of the Sierra Nevada, nitrogen deposition is not believed to be significant in the mid elevation forests. In the south central Sierra Nevada, estimates of deposition are in the range of 2–4 kg ha<sup>-1</sup> yr<sup>-1</sup> (Bytnerowicz et al., 2002; Chorover et al., 1994). However, during prescribed or wildfire events, local NO<sub>x</sub> concentrations can become quite high (Bytnerowicz et al., 2002). With increasing population, fossil fuel use, continued excessive fertilizer use in the San Francisco Bay area and San Joaquin Valley, and increased frequency of anthropogenically-caused fires, we can expect nitrogen deposition to Sierra Nevada ecosystems to become a chronic problem. In areas experiencing moderately high O<sub>3</sub> and above (e.g., > 280 ppm h based on 24 h exposure over the 6 month growing season) such as the southern Sierra Nevada, nitrogen deposition may also be a significant co-stressor. In Sequoia National Forest near Mountain Home, high nitrogen deposition is suspected to cause forest tree responses similar to those found in the Transverse Range in southern California (Fenn et al., Chapter 5, this volume).

The focus of this chapter is western conifer response to oxidant exposure. Most of the examples are based on two yellow pine species, ponderosa and Jeffrey (*Pinus ponderosa* Dougl. Ex Laws. and *Pinus jeffreyi* Grev. & Balf.) responses because these two pines are present on nearly all of the forest plots surveyed for O<sub>3</sub> injury in the Sierra Nevada. Ozone exposure in the Sierra Nevada is greatest at lower elevations, and decreases with increasing elevation and distance from source (Bytnerowicz et al., 2002). Consequently, coniferous species that form lower treeline, such as Coulter (*Pinus coulteri* D. Don) and ponderosa pine, may have greater exposure and injury than species at upper treeline, such as lodgepole pine (*Pinus contorta* Dougl. ex. Loud.), red fir (*Abies magnifica* A. Murr.), and Jeffrey pine. The mid-elevation, Sierran

mixed conifer zone is dominated by white fir (*Abies concolor* (Gord. & Glend.) Lindl. ex Hildebr.), ponderosa and Jeffrey pine, incense cedar (*Libocedrus decurrens* Torr.), and relatively small amounts of sugar pine (*Pinus lambertiana* Dougl.). Unique to the Sierra Nevada are also stands of giant sequoia (*Sequoiadendron giganteum* (Lindl.) Buchholz) locally dominant in “groves.” Of these species, ponderosa and Jeffrey pine are the most sensitive conifers to O<sub>3</sub> exposure (Miller et al., 1983), and as such, are the “bell weathers” for forest health in the Sierran mixed conifer zone.

The interaction of environmental factors, plant phenology (the timing of growth events and birth and mortality of plant parts), physiological status (nutritional or moisture status, and dormant or actively growing within the year), and tree age (interannual differences in resource acquisition and requirements for allocation) all contribute to the complexity of conifer response to O<sub>3</sub> exposure. Of relevance to this book, however, is the translation of the whole tree physiological response to O<sub>3</sub> exposure, in the collection of relatively few morphological (visible) attributes that are used to assess O<sub>3</sub> injury in the field.

The objective of this chapter is to describe seasonal variation in attributes used to assess O<sub>3</sub> injury, under a range of O<sub>3</sub> exposures and nitrogen deposition fluxes typical for the Sierra Nevada. The relationship between O<sub>3</sub> exposure and O<sub>3</sub> uptake, and their influence on the biological response attributes used to assess injury in the field will be discussed. Responses in different microsites (mesic vs. xeric) and in years of significantly different water availability (average or above and 20% below average) will be used to elucidate the effect of drought as an additional environmental stressor. Perhaps the most useful application of this information is to help determine in what month, and in which year to make consistent O<sub>3</sub> injury assessments based on what we know about the effect of environmental factors and temporal variation in attributes.

## 2. Attributes used in O<sub>3</sub> injury assessments of conifers

The attributes used in O<sub>3</sub> injury assessments include foliar chlorotic mottle and or necrosis, both branch (number of needle age classes retained) and within-whorl needle retention, needle length, and proportion of the live crown remaining. Trees used in assessment have already been selected for dominance or co-dominance, and to minimize the effect of pathogens (bark beetle, mistletoe, bud worm).

There are two commonly used assessment methods used in Sierra Nevada forests: O<sub>3</sub> injury index (OII; Miller et al., 1996a) and Forest Pest Management (FPM; Pronos et al., 1978). In the OII assessment, five attributes are utilized for dominant or co-dominant yellow pine: chlorotic mottle, number of whorls retained, needle retention within whorls, needle length, and percent live crown.

The higher the OII score, the more oxidant injury. In the FPM assessment, the first needle age class that exhibits chlorotic mottle for more than one third of that foliage age class is recorded. In this assessment, the lower the score, the more oxidant injury. Arbaugh et al. (1998) has recently described these methods in more detail, as well as their development.

Additional attributes correlated with oxidant exposure have also been reviewed and statistically assessed for significant correlation with level of chlorotic mottle in Grulke and Lee (1997). In that paper, eight attributes were highly correlated with level of chlorotic mottle: foliar nitrogen or chlorophyll content (interchangeable in the multivariate analysis), number of whorls retained, a quantitative measure of within whorl retention, a measure of wood production (basal area increment or branch diameter), spring root carbohydrate content, and distance to the nearest conspecific neighbor as a measure of inferred competition. Foliar antioxidant concentrations and activities have also been clearly correlated with oxidant exposure levels and degree of foliar chlorotic mottle (Tausz et al., 2001). Tissue chemistry, and biochemical attributes (carbohydrates, pigments, antioxidants) are currently not a component of any field assessment of oxidant injury: they are costly and time consuming to assess. However, these as well as other attributes may be chosen for future assessments. Biochemical attributes may be particularly important in assessing response of species that are asymptomatic.

### **3. What is chlorotic mottle?**

Of all the attributes used in O<sub>3</sub> assessment on the many different species in the Sierra Nevada, visible injury to the leaf or needle is the first step of O<sub>3</sub> injury assessment. In yellow pine, the visible injury is chlorotic mottle (Fig. 1; Miller et al., 1963, 1996a). The two most common assessments, FPM and OII survey yellow pine. The FPM method of O<sub>3</sub> assessment is wholly based on the first needle age class with significant chlorotic mottling, functionally greater than 10% on more than a third of the needles of that whorl. Forty percent of the OII score is based on chlorotic mottle of all needle age classes retained (up to 6 years).

Although no quantitative assessment techniques similar to that used for yellow pine have been developed for other species, foliar symptoms may also develop in response to O<sub>3</sub> exposure. In white fir, chlorotic mottling on the needle margins or at the tip or necrosis is apparent with O<sub>3</sub> exposure (Taylor et al., 1980). Seedlings of giant sequoia up to three years old also have chlorotic mottling of needles under current ambient pollutant levels (Miller et al., 1996b). Older giant sequoia trees at ambient or experimentally elevated O<sub>3</sub> levels do not have visible symptoms on foliage (Grulke et al., 1989, 1996). Incense cedar



*Figure 1.* Example of chlorotic mottle development on ponderosa pine on the left. The oldest foliage within a needle age class senesces first, as seen in the previous year whorl in photo on the right. This photo was taken at the end of July in a drought year (1994) at Strawberry Peak, in the San Bernardino Mountains.

is also asymptomatic at ambient  $O_3$  levels. Other pines, such as lodgepole and sugar appear to be more tolerant to  $O_3$  exposure than the yellow pines (Miller et al., 1983), but have similar symptoms: chlorotic mottle that begins near the tip. Most of the pines exhibit some foliar injury at current ambient  $O_3$  levels (approximately 260 ppm h, cumulative  $O_3$  exposure over the 6 month growing season; National Park Service ARD data base) in Sequoia National Park, the most polluted National Park in the country.

Because ponderosa pine is the most sensitive conifer to oxidant exposure, some individuals exhibit visible symptoms even at relatively clean sites in the southern Cascade Mountains in California (Grulke, 1999). There,  $O_3$  concentrations are just slightly elevated over global background levels (38 to 42 ppb per hour averaged over the six month growing season; Grulke, 1999). At this  $O_3$  exposure level (180 ppm h), 1% of the trees exhibited foliar injury symptoms.

Because  $O_3$  concentrations are commonly moderate and above during the growing season in the western United States (Lee and Hogsett, 2001), and because of its high phytotoxicity,  $O_3$  probably has the most negative effect of any air pollutant at ambient levels on tree vigor and growth (Reich, 1987). Once in the substomatal cavity,  $O_3$  reacts with water and or cell membranes or

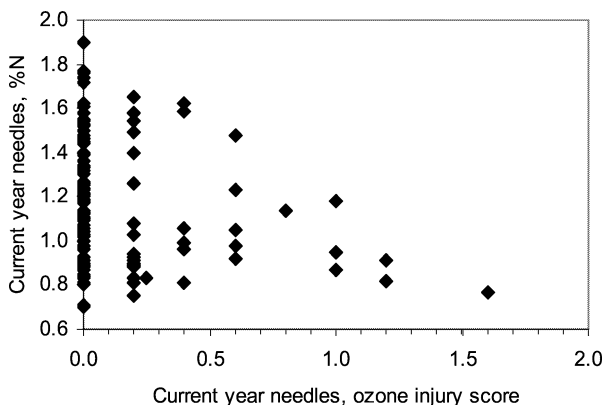


Figure 2. Relationship between chlorotic mottle index and leaf nitrogen content (% dry weight). Samples were taken in an above average precipitation year (1992), and current year foliage was collated for five branches per tree. The chlorotic mottle index corresponds to the following score: 0: no chlorotic mottle; 1: 1–6%; 2: 7–25%; 3: 26–50%; 4: 51–75%; 5: 76–100% chlorotic mottle.

walls to form reactive oxygen species (Polle, 1998). It is not clear if reactive oxygen species are caused by reaction of  $O_3$  and water (Byvoet et al., 1995; Grimes et al., 1983), or by  $O_3$  and constituents of the cell wall and membrane (Mehlhorn et al., 1990). Highly reactive oxygen species disrupt membrane integrity, pigment structure, and oxidize proteins (Mehlhorn et al., 1986) contributing to chlorotic mottle development and or cell death. Reactive oxygen species may also act as a signal to initiate or coordinate other processes such as ethylene production, which induces senescence (Pell et al., 1997; Schraudner et al., 1997). Reactive oxygen species are detoxified by antioxidant systems in the apoplasm, cytoplasm, and chloroplast (Alscher et al., 1997; Heath and Taylor, 1997; De Kok and Tausz, 2001).

Photosynthesis is inhibited by  $O_3$  exposure (Weber et al., 1993): both pigment structure and RUBP carboxylase activity are altered (Schweizer and Arndt, 1990). When the photosynthetic pigments have been damaged, the pigment must be fully broken down (and or new N and Mg must be taken up and transported to the leaf) for pigment to be regenerated (Bjorkman and Demmig-Adams, 1995). The relationship between chlorotic mottle estimates and chlorophyll content or leaf nitrogen content is not precise (Fig. 2). Chlorotic mottle is by definition patchy and on a whole needle basis, nitrogen content on a per leaf basis may not differ between symptomatic and asymptomatic needles. Needles exposed to both  $O_3$  exposure and drought stress had higher nitrogen content than that of controls because significant foliar biomass was lost, and nitrogen may have been translocated out of senescing into remaining needles (Temple and Riechers, 1995). This explains in part for the within-growing season

changes in chlorotic mottle estimates: older needles (generally more symptomatic) within the whorl may be lost through the growing season, and thus the chlorotic mottle estimate decreases for the whole whorl. Higher chlorotic mottle is significantly correlated to lower net assimilation in Jeffrey pine (Patterson and Rundel, 1995).

#### 4. Factors influencing the expression of chlorotic mottle

Perhaps the simplest demonstration of the effect of environmental stressors on the expression of chlorotic mottle can be made by plotting the relationship between chlorotic mottle and cumulative O<sub>3</sub> exposure or uptake for three sites differing in pollutant exposure and between years with average or below-average total annual precipitation (Fig. 3). These sites have been described previously (O<sub>3</sub> exposure, Grulke, 1999; O<sub>3</sub> uptake, Grulke et al., 2002a), and range from roughly 60 (Barton Flats) to 70 (Strawberry Peak), to 80 ppb (Crestline) average O<sub>3</sub> concentration per hour (24 h per day, April 15 through October 15), and low (6–9 kg ha<sup>-1</sup> yr<sup>-1</sup>) to high nitrogen deposition (30–40 kg ha<sup>-1</sup> yr<sup>-1</sup>; Kiefer and Fenn, 1997).

In an average precipitation year, percent chlorotic mottle increased regularly with cumulative O<sub>3</sub> exposure, but the same level of cumulative O<sub>3</sub> exposure did not elicit the same level of chlorotic mottle between sites in that year. In the 20% below average precipitation year, chlorotic mottle developed more quickly at the moderate and moderately high pollution site: the peak expression occurred at the end of June. At both of these sites, there was a significant reduction in chlorotic mottle in the last month of the growing season at the two most polluted sites, and in the last two months of the growing season at the moderate pollution site. Drought stress decreased chlorotic mottle of the whole whorl because the older, more damaged needles within the whorl were lost (Miller et al., 1996b). When cumulative O<sub>3</sub> uptake (calculated by a physiologically based model, or empirically) is used instead of exposure in the relationship, there was no improvement in the relationship with percent chlorotic mottle. These results strongly suggest that interannual comparisons of ozone injury should not be made between years that significantly differ in soil moisture availability (e.g., average vs. below-average precipitation years).

The most polluted site based on cumulative O<sub>3</sub> exposure was also the site of the greatest O<sub>3</sub> uptake by the trees, whether it was a year of average or below-average precipitation. The converse was true for the moderate pollution site, Barton Flats (Grulke et al., 2002a). The maximum chlorotic mottle within the growing season roughly followed the rank order of cumulative O<sub>3</sub> exposure at the three sites (Fig. 4). For example, at the most polluted site, maximum chlorotic mottle was 35% at the most polluted site, 20% at the moderately

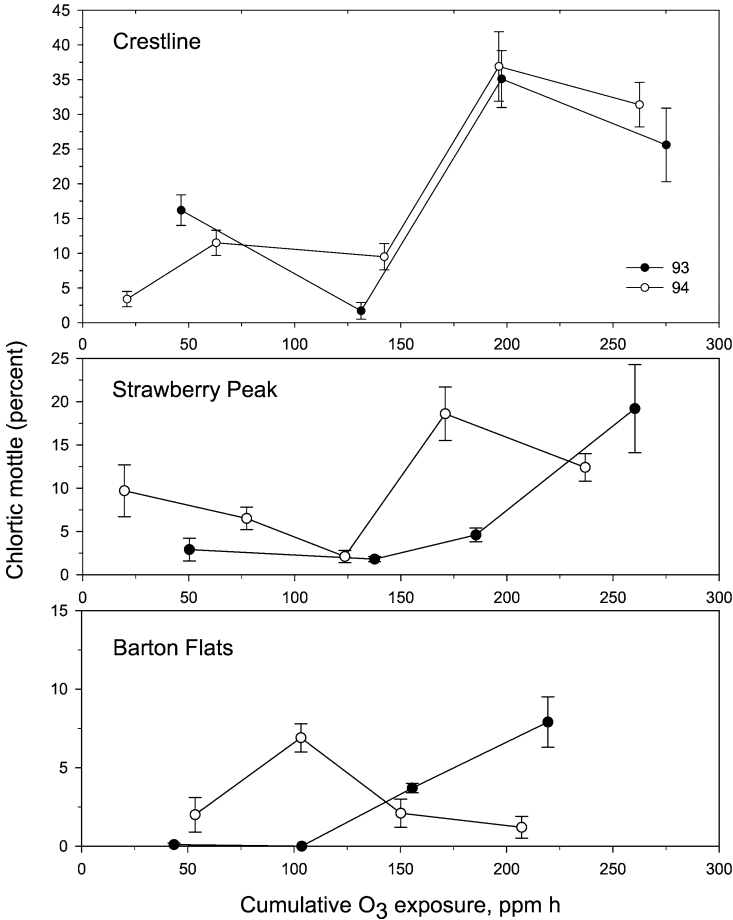


Figure 3. Relationship between chlorotic mottle and cumulative O<sub>3</sub> exposure at three sites in the San Bernardino Mountains (Crestline—highly polluted; Strawberry Peak—moderately high pollution; Barton Flats—moderate pollution) in a year of average (1993) and below-average total annual precipitation (1994). Chlorotic mottle of the previous year only has been plotted because this needle age class has the highest carbon gain in ponderosa pine. Chlorotic mottle was monitored monthly on five branches per tree on twelve trees per site.

high pollution site, and 8% at the moderate pollution site. In the drought year, the seasonal maximum occurred at much lower cumulative O<sub>3</sub> exposure. If the seasonal minimum and maximum percent foliar chlorotic mottle from all sites is plotted with respect to cumulative O<sub>3</sub> exposure, there appears to be a relationship (increase in chlorotic mottle with increase in O<sub>3</sub> exposure), but the relationship is not statistically significant (Adj.  $r^2 = 0.31$ ;  $P = 0.06$ ). A slightly

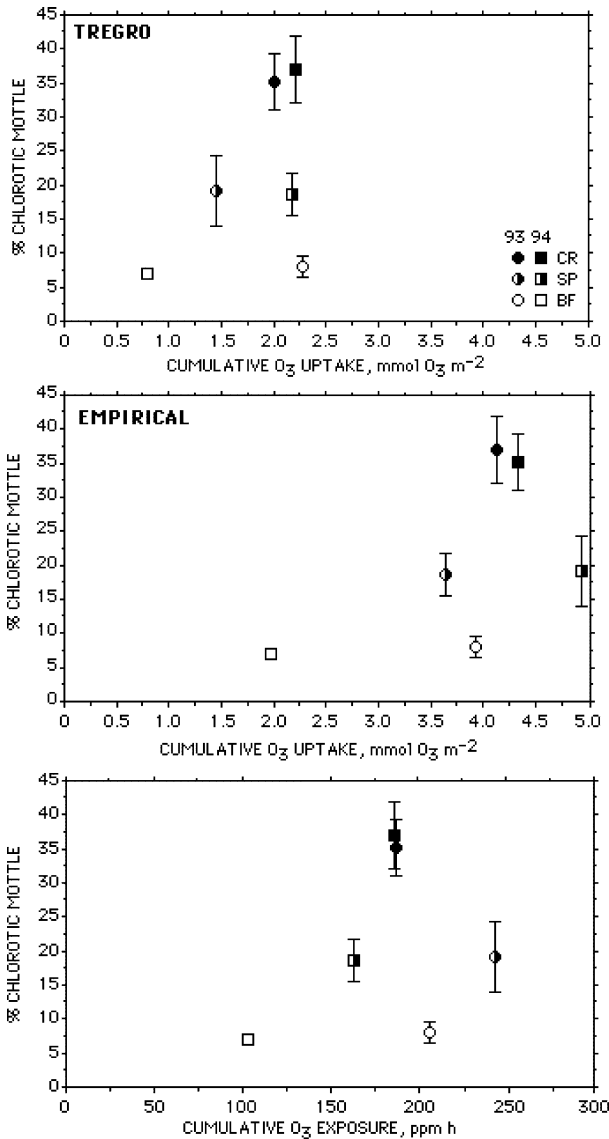


Figure 4. The seasonal maximum chlorotic mottle (from Fig. 3) is plotted for three sites across a pollution gradient with respect to O<sub>3</sub> exposure (lower graph), O<sub>3</sub> uptake (middle graph, as estimated from an empirical model (Grulke et al., 2002a); and upper graph), as estimated from a physiological model, TREGRO; courtesy of C.C. Fan and W.A. Retzlaff). Data in a year of average precipitation (1993) and a drought year (1994) is presented based on a 120 yr record maintained by the San Bernardino County Water District. The percent chlorotic mottle is not well correlated to cumulative O<sub>3</sub> exposure. Cumulative O<sub>3</sub> uptake, whether empirically or physiologically modeled, was a better predictor of response.

better fit was obtained with cumulative O<sub>3</sub> uptake (Adj.  $r^2 = 0.35$ ;  $P = 0.04$ ). However, the cumulative O<sub>3</sub> exposure at the time of maximum seasonal expression was not significantly correlated to the maximum percent chlorotic mottle. Cumulative O<sub>3</sub> uptake as calculated by a physiological model, TRE-GRO, had the same predictive capability as that of statistically calculated O<sub>3</sub> uptake (Adj.  $r^2 = 0.34$ ;  $P = 0.04$ ; CC Fan and WA Retzlaff, unpubl. simulation for ponderosa pine using the same data set). Site O<sub>3</sub> exposure alone does not wholly explain the physiological response and may interact with environmental factors in unexpected ways. However, a significant correlation coefficient was obtained between a chlorotic mottle score and several O<sub>3</sub> exposure indices (7 and 24 h SUM00, and 7 and 24 h SUM06; Salardino and Carroll, 1998) of forest stands distributed along the Sierra Nevada.

Foliar oxidant injury usually accelerates only after the upper soil horizons have dried out (Grulke and Balduman, 1999). Although foliar N and chlorophyll at high nitrogen deposition and O<sub>3</sub> exposure sites is high early in the summer, foliar N and chlorophyll content decreases with increasing O<sub>3</sub> exposure after mid summer (Grulke, 1999). Cumulative O<sub>3</sub> exposure, high radiation, and drying of the upper soil horizons (insufficient uptake of N to replenish damaged pigments) perhaps all contribute to the expression of chlorotic mottle. Cumulative O<sub>3</sub> exposure early in the summer is significant (Grulke et al., 2002a), but upper soil horizons are rich in nitrogen and fine root mass is high until they dry out, even in a year of average precipitation, in mid to late July. When fine roots (and associated mycorrhizae and microbial communities) die off in near-surface soil horizons, trees rely on deep roots for water (Hubbert et al., 2001). Even at sites where the ground water is also rich in nitrogen (Fenn and Poth, 1999), chlorotic mottle still develops. Either deep roots (and associated mycorrhizae) are not as effective at nitrogen uptake, or oxidant damage is too great for reparation activities to keep up with N uptake from the pedosphere. In the autumn with favorable soil temperatures and moisture permitting, fine roots and mycorrhizae grow in the near surface soil horizons, and chlorotic mottle can significantly decrease (Fig. 5).

Because OII integrates foliar response of up to six needle age classes, theoretically it should not be correlated to cumulative O<sub>3</sub> exposure in any one year of assessment unless site cumulative O<sub>3</sub> exposure is fairly consistent from year to year. In order to generate a meaningful value for site O<sub>3</sub> exposure, O<sub>3</sub> exposure should be accumulated over the average number of years that needles are retained on the branches, as described in Arbaugh et al. (1998) and tested in Salardino and Carroll (1998). The OII scoring system works well to rank significant differences in site to site injury level. The FPM is also a robust scoring system that works well to coarsely rank stand injury under a wide range of environmental co-stressors. Integrating cumulative O<sub>3</sub> exposure for this score is even more problematic than for OII because chlorotic mottle is not integrated

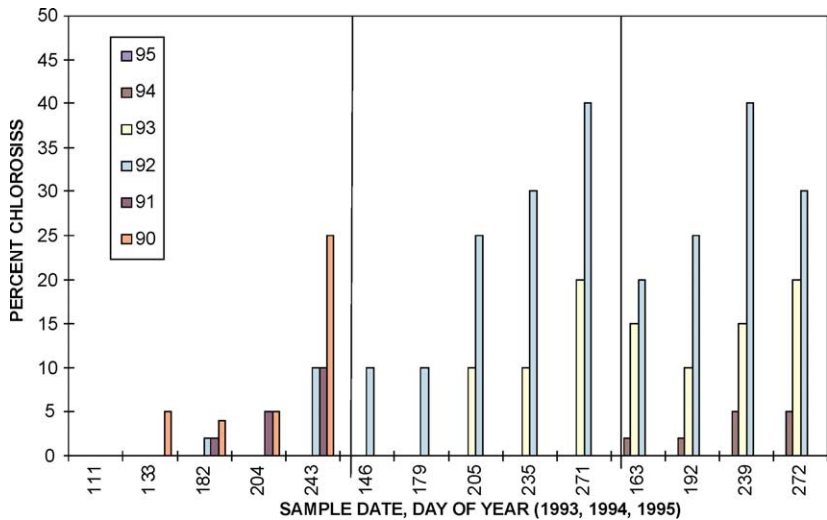


Figure 5. Typical phenology of chlorotic mottle in each needle age class, given for one branch, over the course of three years, in the San Bernardino Mountains (Grulke, unpubl. data).

over all needle age classes, but is based on the needle age where chlorotic mottle is first significantly expressed. In both scoring systems, chlorotic mottling is modified by the degree of overwinter reparation of damaged pigments (Fig. 5).

Perhaps the most effective use of the FPM and OII score would be to conduct canopy assessments in average or above-average precipitation years, preceded by at least one average or above precipitation year to best rank injury in widely ranging geographic sites. Assessment in a below-average precipitation year, or of stands with a disproportionate number of trees in xeric microsites could significantly decrease the OII score, or increase the FPM score (Fig. 6) if assessments were made on the same day of year. Both scores reflect a microsite difference in  $O_3$  uptake, based on both leaf-level and canopy-level measurements (Grulke et al., in press). Trees in mesic sites had 20% greater seasonal  $O_3$  uptake than Jeffrey pine trees in xeric sites. Choosing trees in consistently mesic microsites should increase the chance of a relationship between site  $O_3$  exposure and foliar injury development. Assessing canopy injury at the same phenological stage (such as near or at the end of needle elongation, correlated with upper soil horizons drying to < 4% soil moisture; Grulke et al., 1998, 2001) would significantly aid in interannual comparisons of foliar injury.

The level of chlorotic mottle could differ between sites with the same cumulative  $O_3$  exposures because of associated nitrogen deposition at these sites. Greater nitrogen availability could aid repair of damaged pigments, but it could also increase stomatal conductance (via stimulated photosynthe-

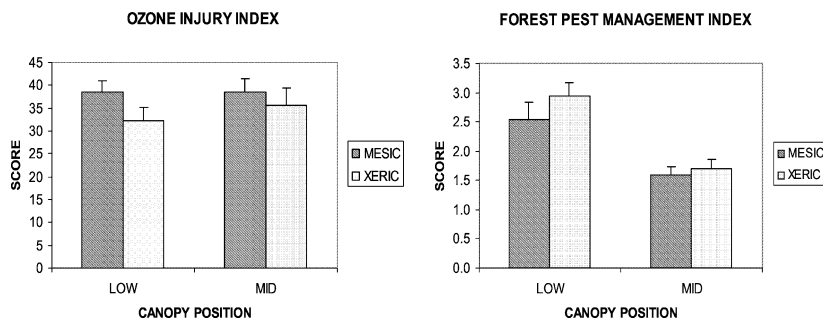


Figure 6. OII and FPM scoring for Jeffrey pine trees in mesic and xeric microsites, at lower and mid canopy. The OII score is much less sensitive to canopy position than FPM. Mesic trees had poorer health as indicated from both the OII (higher score, more injury) and the FPM (lower score, more injury) assessment techniques. Leaf-level and canopy-level measures of gas exchange indicate that trees in mesic sites had 20% greater seasonal  $O_3$  uptake than trees in xeric sites (Grulke et al., in press) given the same site  $O_3$  exposure.

sis), and thus  $O_3$  uptake. From a biochemical point of view, nitrogen-limited trees had greater antioxidant concentration than that of controls (Polle, 1998). Conversely, excess nitrogen in fertilizer manipulations increased secondary metabolites with antioxidizing capacity (Pell et al., 1995).

Experimental applications of nitrogen to trees in consistently mesic or xeric microsites on the same slope may help determine the direction of change expected with concurrent environmental stressors ( $O_3$  exposure, drought) and enhancers (nitrogen deposition). After three years of experimental amendments of nitrogen availability ( $+50 \text{ kg ha}^{-1} \text{ yr}^{-1}$ ), mid canopy chlorotic mottle decreased, foliar nitrogen increased slightly, foliar carbon decreased, and foliar C:N decreased relative to control trees in mesic microsites for mature Jeffrey pine in Sequoia National Park (Fig. 7). The opposite trends occurred in midcanopy foliage of trees in xeric microsites. In a similar experiment, N-fertilized ( $+50 \text{ kg ha}^{-1} \text{ yr}^{-1}$ ) mature yellow pine at a dry site had lower bole growth than that of controls (Fenn, unpubl. data). The two experiments had similar background  $O_3$  exposures, approximately 260 ppm h accumulated over the growing season. At a high pollution site (approximately 320 ppm h seasonal  $O_3$  exposure,  $30\text{--}40 \text{ kg ha}^{-1} \text{ yr}^{-1}$  background N deposition) with more moisture availability, additional N amendments ( $+50 \text{ kg ha}^{-1} \text{ yr}^{-1}$ ) increased bole growth. In dry sites, perhaps N amendments decrease fine root mass and pedosphere N uptake. In mesic sites, fine root (+ mycorrhizae) may be able to take up and transfer more N to the foliage. Perhaps foliar carbon content of fertilized trees is lower in mesic sites because of within-tree allocation to bole growth (or other tissue), N-stimulated foliar growth reduced specific leaf weight, or increased respiration due to greater  $O_3$  exposure in this site.

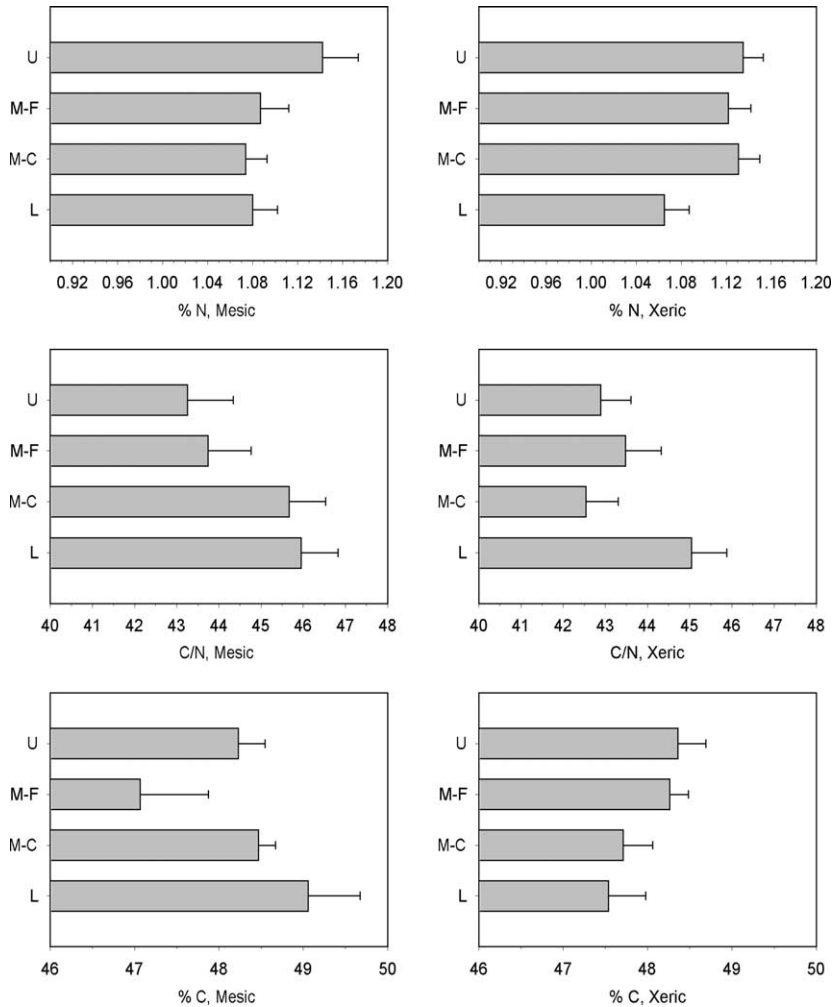


Figure 7. Within canopy variability in carbon, nitrogen, and C:N ratio in Jeffrey pine, and effect of 3 years of nitrogen amendments on mid canopy tissue chemistry. Each bar represents the average and  $\pm 1$  S.D. of foliage from 32 trees. Within canopy foliar chemistry differed between trees in mesic and xeric microsites. Nitrogen amendments increased foliar nitrogen, decreased carbon, and decreased C:N of mid canopy foliage on mesic trees. It had the opposite effect on tissue chemistry of xeric trees. Trees were assigned to different microsites based on topographic position, proximity to perennial surface water, and ability of mesic site trees to increase bole growth in average and above precipitation years (Grulke et al., in press).

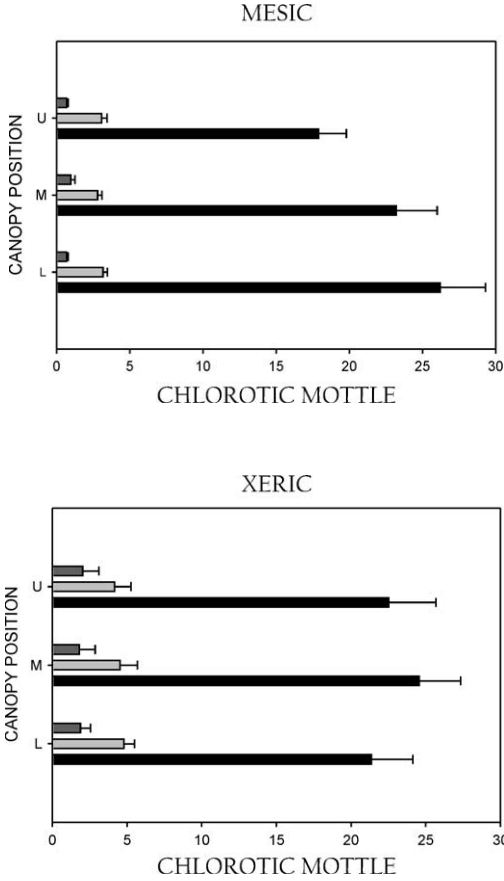


Figure 8. Within-canopy variability in foliar chlorotic mottle in Jeffrey pine, Sequoia National Park. Each bar at each canopy position (U = upper; M = middle; L = lower) represents chlorotic mottle of current (dark grey), previous year (light grey), and four year old needles (black). Bar length represents the average and  $\pm 1$  S.D. of 32 trees in each microsite (same trees as presented in Fig. 7).

The level of chlorotic mottle could differ between sites with the same cumulative  $O_3$  exposures because of differences in drought stress. Jeffrey pine in xeric microsites had 20% lower  $O_3$  uptake than mesic microsite trees, but chlorotic mottle was greater at all canopy levels in current and previous year needles, and in four year old needles in the upper canopy (Fig. 8; Grulke et al., in press). This may be a field example of free radical production within the chloroplast due to drought stress, lower stomatal conductance, lower  $C_i$ , and

high radiation loads (Tausz et al., 2001; Foyer et al., 1994). These trees had greater antioxidant concentrations relative to mesic microsites in June, prior to the onset of seasonal drought stress (Tausz et al., 2002). In late summer, drought stress and O<sub>3</sub> uptake combined to decrease carbon acquisition in ponderosa pine (Grulke et al., 2002b). In simulations with white fir, a 25% reduction in annual precipitation at moderate O<sub>3</sub> levels was protective against injury as assessed by tree biomass (Retzlaff et al., 2000). Drought stress imposed at greater levels in combination with moderate O<sub>3</sub> exposure was deleterious to carbon acquisition in white fir.

### 5. Factors influencing needle retention (branch-level; whorl-level)

There are two types of needle retention measurements of interest in O<sub>3</sub> injury assessments: the number of live needle age classes on the branch, and the within needle age class (whorl) retention. The former is represented with a simple count, and the latter is either estimated by thirds (Miller et al., 1996a) or measured ( $[\text{length of the branch with live needles}] / [\text{branch elongation growth}]$ ) in a given year; Grulke and Balduman, 1999). In OII, the branch-level needle retention (the number of needle age classes or whorls retained on the branch), and the within-whorl needle retention is 40% of the O<sub>3</sub> injury score. Long-lived foliage acts as a storage location for nitrogen, but with a carbon cost for maintenance (Chapin, 1980). Ozone exposure accelerates needle senescence (Heath and Taylor, 1997; Pell and Dann, 1991), with the net result that N can be translocated out of senescing tissue to replace damaged photosynthetic pigments. Ozone exposure can also increase foliar respiration due to reparation: it may become too costly to maintain older needles with lower assimilation rates.

Nitrogen deposition, positively correlated with O<sub>3</sub> exposure, also accelerates needle loss (Gower et al., 1993), but via a different mechanism. Nitrogen amendments increase foliar nitrogen contents, and older leaves may have redundant nitrogen with too high carbon cost for maintenance. Trees with nitrogen amendments in Sequoia National Park had higher foliar nitrogen content, and fewer needle age classes relative to control trees in mesic microsites (Fig. 7; Grulke, unpubl. data).

The FPM score is described as the youngest needle age class with significant chlorotic mottle development. However, the whorl number and the level of chlorotic mottle may co-vary, as suggested by the ratio of chlorotic mottle and foliar retention as a new index ("CM/FR," Salardino and Carroll, 1998). Ponderosa pine trees with known allelic and genotypic diversity at a site with both high O<sub>3</sub> and nitrogen deposition illustrate the interaction between chlorotic

mottle and needle retention (Staszak et al., unpublished data; Prus-Glowacki et al., 1999). Ponderosa pine with a genetic signature indicating sensitivity to O<sub>3</sub> had two strategies: (1) many needle age classes retained, but high chlorotic mottle across all needle age classes (e.g., FPM score, 0); and (2) few needle age classes retained, but little chlorotic mottle (e.g., FPM score, 3). Also, ponderosa pine with a tolerant O<sub>3</sub> genetic signature less than 2 m from a conspecific tree had morphological and tissue chemistry attributes similar to a sensitive tree (lower foliar nitrogen, higher chlorotic mottle, fewer needle age classes) than typical for a “tolerant” tree.

Drought stress also reduces both the number of needle age classes retained, and retention within a needle age class. The response of both types of needle retention is illustrated in Fig. 9 for four sites differing in pollutant exposure. The number of needle age classes retained significantly declined with increasing pollutant exposure, as has been described previously by both Pronos et al. (1978) and Miller et al. (1996a). At the relatively clean atmosphere site, there was little difference in within-whorl needle retention between an average and below-average precipitation year. In the southern Californian sites within increasing pollutant exposure, there was significant loss of both whorls and within-whorl needle retention in a drought year. Drought stress acts synergistically with O<sub>3</sub> exposure to promote premature senescence of foliage. Both nitrogen (from reduced root mass and uptake) and carbon (increasingly lower net carbon balance with reduced stomatal aperture, lower assimilation, and constant respiratory losses) may be limiting in a drought year to facilitate this response.

The quantitative measure of within-whorl retention is a better measure of needle retention than the estimated needle retention score of Miller et al. (1996a) for ponderosa pine. Ponderosa pine loses needles regularly from the lower or oldest needles to the upper or younger needles within a whorl. However, Miller et al. (1996a) needle retention score is a better measure for Jeffrey pine because needles are lost erratically within the whorl.

Male cone scars can sometimes be mistaken for needle loss within the whorl: their scars would reduce estimated needle retention and increase the OII score (implying greater injury). Each male cone develops at the expense of fascicle production (for lodgepole pine, McDick et al., 1990), thus needle mass will necessarily be lower in whorls that bear them. Female cones develop from terminal buds, thus form at the expense of branch growth (for Monterey pine, *Pinus radiata* D. Don, Cremer, 1992). Both male and female cone production accounted for 5% of total aboveground production in this species, but this would not be detected as a reduction in crown mass in a field assessment.

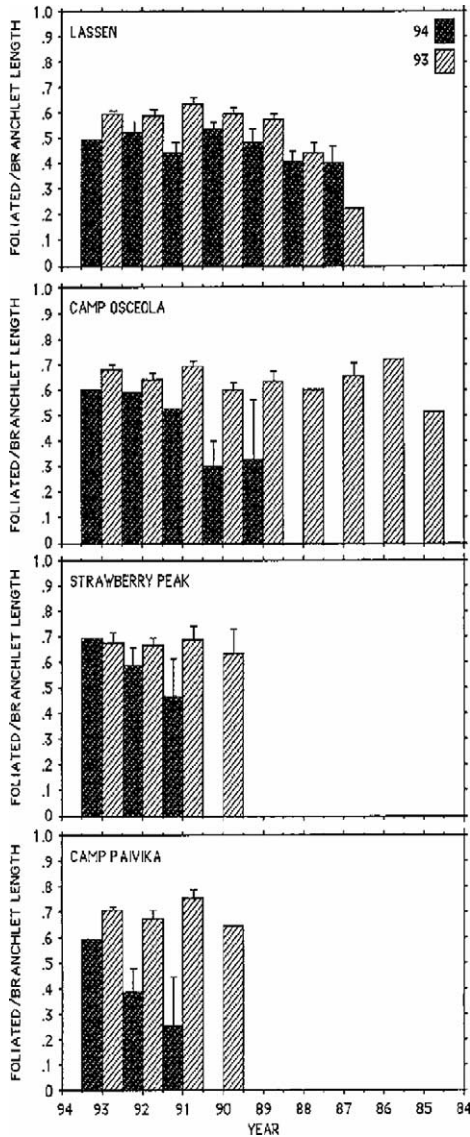


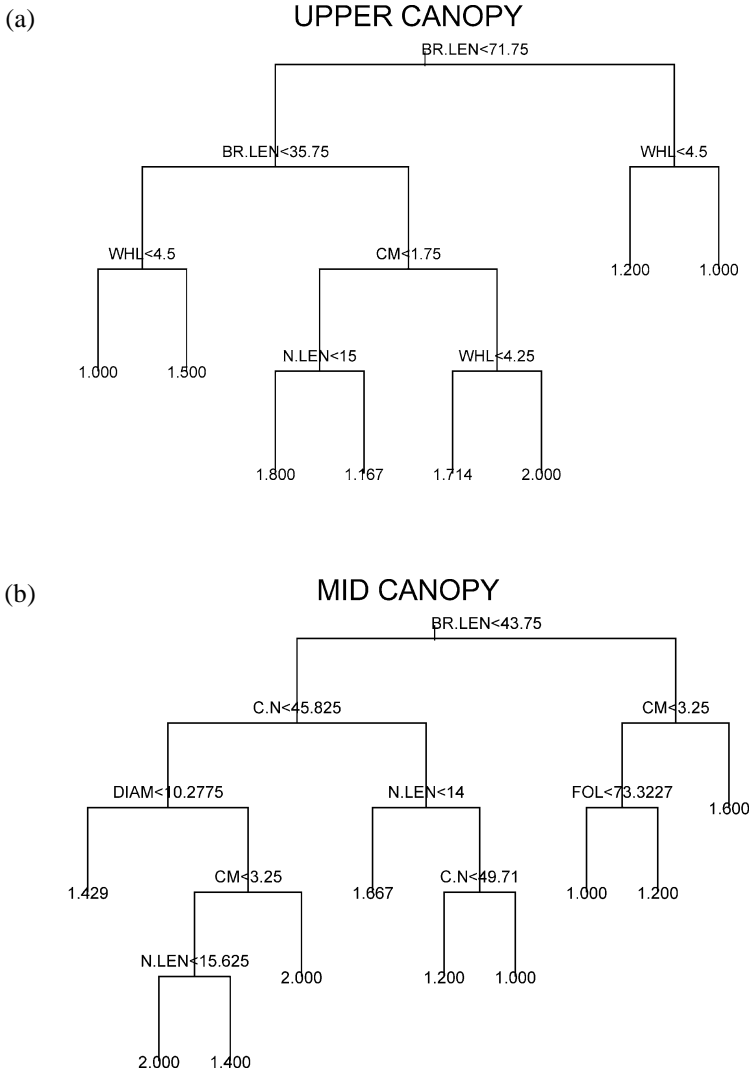
Figure 9. Example of both branch-level and whorl-level needle retention in a year of above-average precipitation (1993) and below average precipitation (1994) (from Grulke and Balduman, 1999). Within-whorl retention is a quantitative measure constructed from the [length of the branchlet with green needles retained]/[total branchlet length]. In this case, the foliated proportion of the branch is calculated for each branch age class, not the entire branch as in Miller et al. (1996a). Reprinted courtesy of Kluwer Academic Publishers B.V., Water, Air, and Soil Pollution, Volume 116, Figure 1, p. 242.

**6. Within-canopy variability in O<sub>3</sub> injury expression**

Both OII and FPM assessments are generally conducted in lower canopy due to ease of access for extensive surveys. To illustrate the difference in expression in O<sub>3</sub> injury within the canopy, morphological and tissue chemistry attributes in three canopy positions of 30 trees each in mesic and xeric microsites were measured. The following attributes were used in the multivariate analysis:

*Table 1.* Summary statistics for upper, mid, and lower canopy in mesic and xeric microsites. Attributes include number of whorls retained, chlorotic mottle (%), needle length (mm), branch length (mm), proportion of the branch foliated (%), branch diameter (mm), and foliar carbon (%) nitrogen (%), and C : N. Data is presented for previous year tissue only. A two-way nested ANOVA was used to test for significance between canopy position (CAN POS), microsite (SITE), and their interaction.

Attribute		Upper	Mid	Lower	Can Pos <i>P</i> =	Site <i>P</i> =	A × B <i>P</i> =																																																																																												
# WHRLS	M	5.0 (0.2)	4.6 (0.2)	4.5 (0.2)	0.050	0.233	0.680																																																																																												
	X	4.7 (0.2)	4.9 (0.2)	4.7 (0.1)				% Chlor	M	3.1 (0.4)	2.8 (0.3)	3.2 (0.3)	0.836	0.007	0.874	X	4.2 (0.4)	4.5 (1.2)	4.8 (0.7)	N length	M	15.6 (0.4)	15.7 (0.4)	15.7 (0.6)	0.286	0.012	0.209	X	15.5 (0.4)	14.8 (0.3)	14.0 (0.3)	BR length	M	65 (5)	41 (3)	34 (2)	0.0001	0.0003	0.242	X	49 (3)	35 (2)	27 (2)	% Foliated	M	75 (2)	68 (1)	64 (2)	0.001	0.014	0.021	X	73 (2)	73 (2)	71 (2)	BR diam	M	14.1 (0.5)	11.6 (0.3)	10.4 (0.3)	0.0001	0.057	0.778	X	14.4 (0.5)	12.4 (0.4)	11.2 (0.3)	% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)	% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442
% Chlor	M	3.1 (0.4)	2.8 (0.3)	3.2 (0.3)	0.836	0.007	0.874																																																																																												
	X	4.2 (0.4)	4.5 (1.2)	4.8 (0.7)				N length	M	15.6 (0.4)	15.7 (0.4)	15.7 (0.6)	0.286	0.012	0.209	X	15.5 (0.4)	14.8 (0.3)	14.0 (0.3)	BR length	M	65 (5)	41 (3)	34 (2)	0.0001	0.0003	0.242	X	49 (3)	35 (2)	27 (2)	% Foliated	M	75 (2)	68 (1)	64 (2)	0.001	0.014	0.021	X	73 (2)	73 (2)	71 (2)	BR diam	M	14.1 (0.5)	11.6 (0.3)	10.4 (0.3)	0.0001	0.057	0.778	X	14.4 (0.5)	12.4 (0.4)	11.2 (0.3)	% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)	% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)								
N length	M	15.6 (0.4)	15.7 (0.4)	15.7 (0.6)	0.286	0.012	0.209																																																																																												
	X	15.5 (0.4)	14.8 (0.3)	14.0 (0.3)				BR length	M	65 (5)	41 (3)	34 (2)	0.0001	0.0003	0.242	X	49 (3)	35 (2)	27 (2)	% Foliated	M	75 (2)	68 (1)	64 (2)	0.001	0.014	0.021	X	73 (2)	73 (2)	71 (2)	BR diam	M	14.1 (0.5)	11.6 (0.3)	10.4 (0.3)	0.0001	0.057	0.778	X	14.4 (0.5)	12.4 (0.4)	11.2 (0.3)	% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)	% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																				
BR length	M	65 (5)	41 (3)	34 (2)	0.0001	0.0003	0.242																																																																																												
	X	49 (3)	35 (2)	27 (2)				% Foliated	M	75 (2)	68 (1)	64 (2)	0.001	0.014	0.021	X	73 (2)	73 (2)	71 (2)	BR diam	M	14.1 (0.5)	11.6 (0.3)	10.4 (0.3)	0.0001	0.057	0.778	X	14.4 (0.5)	12.4 (0.4)	11.2 (0.3)	% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)	% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																																
% Foliated	M	75 (2)	68 (1)	64 (2)	0.001	0.014	0.021																																																																																												
	X	73 (2)	73 (2)	71 (2)				BR diam	M	14.1 (0.5)	11.6 (0.3)	10.4 (0.3)	0.0001	0.057	0.778	X	14.4 (0.5)	12.4 (0.4)	11.2 (0.3)	% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)	% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																																												
BR diam	M	14.1 (0.5)	11.6 (0.3)	10.4 (0.3)	0.0001	0.057	0.778																																																																																												
	X	14.4 (0.5)	12.4 (0.4)	11.2 (0.3)				% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)	% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																																																								
% C	M	48.3 (0.3)	48.4 (0.2)	49.0 (0.6)	0.817	0.032	0.165																																																																																												
	X	48.4 (0.3)	47.7 (0.4)	47.5 (0.4)				% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)	C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																																																																				
% N	M	1.13 (0.03)	1.08 (0.02)	1.08 (0.02)	0.032	0.558	0.418																																																																																												
	X	1.14 (0.02)	1.13 (0.02)	1.06 (0.02)				C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																																																																																
C : N	M	43.6 (1.0)	45.5 (0.9)	45.9 (0.8)	0.045	0.050	0.442																																																																																												
	X	42.9 (0.7)	42.6 (0.8)	45.0 (0.8)																																																																																															



*Figure 10.* Tree models of morphological and tissue chemistry attributes that significantly contribute to distinguishing between trees in mesic and xeric microsites in the upper, mid, and lower third of the canopy. Numbers at the base of the dendritic line indicate the proportion of trees in mesic (1.0) or xeric (2.0) microsites that satisfy the series of criteria in higher level branches. Foliage or branch tissue of the previous year was measured in thirty Jeffrey pine trees in each microsite, in Sequoia National Park in mid August, 2000. BR LEN = branch length; CM = chlorotic mottle; C.N = C : N ratio in needles; DIAM = branch diameter; FOL = proportion of branchlet with needles retained (see Grulke and Balduman, 1999); N LEN = needle length; WHL = number of live needle age classes retained. (*Continued on next page*)

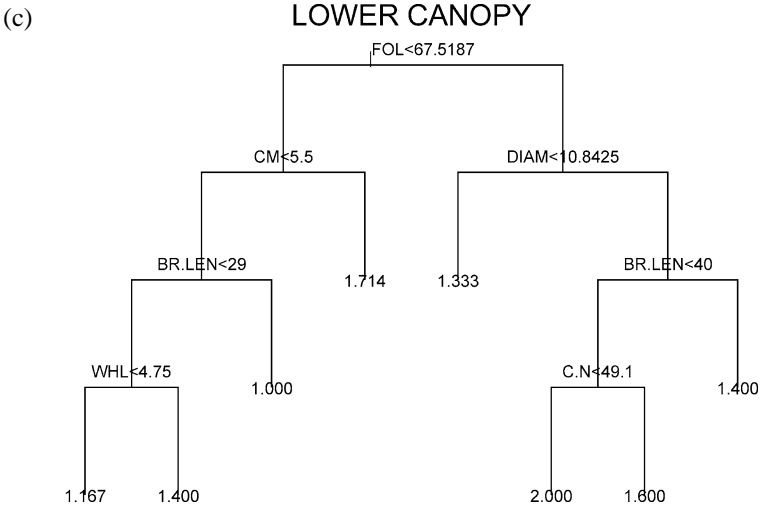


Figure 10. (Continued)

chlorotic mottle, % foliar nitrogen and carbon, foliar C : N, average number of needle age classes retained, proportion of the branch foliated, and branchlet diameter. These attributes were chosen based on the statistical analysis of ponderosa pine described in Grulke and Lee (1997), and subsequent application to Jeffrey pine (Grulke et al., in press). Table 1 provides summary statistics for these attributes for upper, mid, and lower canopy of mesic and xeric trees (prior to multivariate grouping).

In contrast to that reported by Muir and Armentano (1988) for ponderosa pine, needle retention (number of whorls retained on the branch and within-whorl foliar retention) in the lower canopy was significantly lower than that in the upper canopy of Jeffrey pine, regardless of microsite. Less foliage in mid-canopy in mesic microsities may be an indication of greater O<sub>3</sub> uptake in this microsite. The greatest portion of the canopy foliar mass is in the middle third of the tree, followed by the upper canopy, then the lower canopy in ponderosa pine (from whole trees harvested in Grulke and Balduman, 1999). Foliar loss in mid canopy due to oxidant injury, or oxidant and drought stress, is perhaps more critical to whole tree carbon and nitrogen balance than foliar loss in lower canopy. Except for the number of whorls retained on the branch and % foliar N, all other attributes significantly differed between microsities. Except for % chlorotic mottle, needle length, and % foliar carbon, all other attributes measured significantly differed with canopy position.

A tree model (Fig. 10) was used to illustrate which attributes, at what values differentiated the two groups of trees (1.0 = all mesic trees; 2.0 = all

xeric trees; values between 1 and 2 indicate the relative proportion of trees in the two microsites) at each canopy level. For example, the average number of whorls retained was not used to differentiate groups in the mid canopy, but was significant in the upper canopy. Whorl number was marginally useful in lower canopy in differentiating groups. Some attributes (the proportion of the branch foliated; foliar N and C content) significantly assisted in differentiating between groups of trees at all canopy levels.

Divisive clustering (S-Plus 2000; figures not presented) was applied to sort trees into different groups according to the values of the same attributes (used in the tree model above) in each canopy position. This multivariate technique yielded the best separation of microsites, with the highest cluster coefficient. However, the groups were robust: other multivariate analyses yielded the same end groups. Many different combinations of attributes were used to optimize the resulting cluster coefficient. Although some of the attributes could have been excluded for one or more of the canopy positions, a consistent approach was used across all three canopy levels.

Clustering analysis applied to the three canopy layers differed in the number of groups identified (three for lower and upper canopy, and two for mid canopy measurements). The first group largely consisted of mesic site trees at all canopy levels, and the second group largely consisted of xeric site trees at all canopy levels suggesting whole tree differences in the expression of these attributes in the two microsites. The third group consisted of a relatively few number of trees (2 out of 3 trees were mesic in the third group of the upper canopy; 5 out of 6 trees were xeric in the third group of the lower canopy). In Group 1, these attributes misclassified 50% of the trees when measured in upper canopy, 15% of the trees when attributes were measured in mid canopy, and 20% of the trees when measured in the lower canopy. If the upper canopy acts as a sink for resources, attributes measured there would be less effective at indicating microsite location. In Group 2, these attributes misclassified 40% of the trees when measured in the upper canopy, 65% of the trees when measured in mid canopy, and 60% of the trees when measured in the lower canopy. Trees in mesic microsites were more consistent in expression of attributes correlated with O<sub>3</sub> injury in yellow pine.

## 7. Conclusions

The O<sub>3</sub> injury assessments that have been developed for yellow pine are robust and were designed for a coarse assignment of forest health. They do well at this and scores reflect O<sub>3</sub> exposure regimes averaged over many years. Of the many known physiological responses to O<sub>3</sub> exposure in conifers, only a few attributes are used to integrate the whole tree response to the stressor. The

intent of this chapter was to review responses of foliar chlorotic mottle and needle retention, the key attributes relevant to field assessments of O<sub>3</sub> injury. Correlative and manipulative examples of foliar chlorotic mottle and needle retention response to (1) O<sub>3</sub> exposure and uptake, (2) nitrogen deposition frequently associated with O<sub>3</sub>, and (3) drought stress in years of significantly below-average annual totals help to describe the range of field responses expected. Understanding sources of variation in the key O<sub>3</sub> injury assessment attributes may improve between-observer, and between-year assessments, and may help interpreting historical data sets.

Several recommendations are offered for future field assessments of O<sub>3</sub> injury. Because Jeffrey pine has lower stomatal conductance, greater antioxidant defense, and differing patterns of loss within a needle age class, scores should be reported separately for the two species of yellow pine at any one site. Interannual differences in the development (and loss) of chlorotic mottle are significant: the peak foliar injury may appear two months earlier in a drought year relative to a year of average precipitation, although the maximum level of foliar injury developed in a given site appears to roughly correlate with interannual, average site O<sub>3</sub> exposure. The differences reported in leaf- and canopy-level gas exchange and thus O<sub>3</sub> uptake for trees in mesic and xeric microsites, the manifestation of O<sub>3</sub> uptake in chlorotic mottle, tissue chemistry, and needle retention suggests strongly for choosing trees in consistently mesic microsites for consecutive, interannual comparisons, or for restricting repeated assessments to years of  $\pm 10\%$  average precipitation years. Due to reduced foliar production within whorls with male cones, these branches should be avoided in field assessments. Although lower canopy is more easily accessible for large surveys, it is the least representative of the canopy. Future field surveys should be based in the mid canopy, allowing the capability of remote sensing to be linked to ground based measures of O<sub>3</sub> injury.

Ozone injury assessments have been carefully developed for yellow pine, and it is possible that this approach can be applied to both sugar and lodgepole pine because despite their relative tolerance to O<sub>3</sub>, both exhibit chlorotic mottling on their needles. Ozone injury assessments will probably never be developed for giant sequoia, white fir, or incense cedar because visible foliar symptoms do not develop to any appreciable extent. Although subtle physiological responses to O<sub>3</sub> exposure can be demonstrated in white fir (Retzlaff et al., 2000) and giant sequoia (Grulke et al., 1989, 1996), the physiological response is not translated to a consistent set of morphological responses unique to O<sub>3</sub> exposure. Without the definitive chlorotic mottle that Miller et al. (1963) correlated to O<sub>3</sub> exposure, the changes in growth and needle retention observed cannot be conclusively assigned to the stressor in a field survey. Oxidative stress can, however, be supported on asymptomatic species using biochemical techniques.

## Acknowledgments

This chapter largely summarizes research supported over the last decade by the Terrestrial Ozone Program, US Environmental Protection Agency, PRIMENET (a joint NPS/EPA program), and the Air Quality Division of the National Park Service. The views presented here have not been reviewed or specifically condoned by either agency. Special thanks to Paul Miller, for introducing me to O<sub>3</sub> injury effects, and Annie Esperanza, who has provided logistical support and field knowledge for much of this work.

## References

- Alscher, R.G., Donahue, J.L., Cramer, C.L., 1997. Reactive oxygen species and antioxidants: relationships in green cells. *Physiol. Plant.* 100, 224–233.
- Arbaugh, M.J., Miller, P.R., Carroll, J.J., Takemoto, B., Procter, T., 1998. Relationships of ozone exposure to pine injury in the Sierra Nevada and San Bernardino Mountains of California, USA. *Environ. Pollut.* 101, 291–301.
- Bjorkman, O., Demmig-Adams, B., 1995. Regulation of photosynthetic light energy capture, conversion, and dissipation in leaves. In: Schulze, E.-D., Caldwell, M.M. (Eds.), *Ecophysiology of Photosynthesis*. Springer-Verlag, New York, pp. 17–48.
- Bytnerowicz, A., Grulke, N.E., 1992. Physiological effects of air pollutants on western trees. In: Olson, R.K., Binkley, D., Bohm, M. (Eds.), *The Response of Western Forests to Air Pollution*. In: *Ecological Studies Series*, Vol. 97. Springer-Verlag, New York, pp. 183–234.
- Bytnerowicz, A., Fenn, M., Glaubig, R., 1996. Dry deposition of nitrogen and sulfur to forest canopies at three plots (calculated on the basis of foliage rinsing, internal uptake of gaseous pollutants, and estimates of deposition to ground). In: Miller, P.R., Chow, J., Watson, J.G. (Eds.), *Assessment of Acidic Deposition and Ozone Effects on Conifer Forest in the San Bernardino Mountains*. Final Report, Contract No. A032-180, California Air Resources Board, Research Division, 2020 L Street, Sacramento, CA 95814, 4-1, 4-75.
- Bytnerowicz, A., Padgett, P., Percy, K., Krywult, M., Riechers, G., Hom, J., 1999. Direct effects of nitric acid on forest trees. In: Miller, P.R., McBride, J.R. (Eds.), *Oxidant Air Pollution Impacts in the Montane Forests of Southern California: A Case Study of the San Bernardino Mountains*. In: *Ecological Studies*, Vol. 134. Springer-Verlag, New York, pp. 270–287.
- Bytnerowicz, A., Tausz, M., Alonso, R., Jones, D., Johnson, R., Grulke, N., 2002. Summer-time distribution of air pollutants in Sequoia National Park, California. *Environ. Pollut.* 118, 187–203.
- Byvoet, P., Balis, J.U., Shelley, S.A., Montgomery, M.R., Barber, M.J., 1995. Detection of hydroxyl radicals upon interaction of ozone with aqueous media or extracellular surfactant: the role of trace iron. *Arch. Biochem. Biophys.* 319, 464–469.
- Chapin, F.S., 1980. The mineral nutrition of wild plants. *Annu. Rev. Ecol. System.* 11, 233–260.
- Chorover, J., Vitousek, P.M., Everson, D.A., Esperanza, A.M., Turner, D., 1994. Solution chemistry profiles of mixed conifer forests before and after fire. *Biogeochemistry* 26, 115–144.
- Cremer, K.W., 1992. Relations between reproductive growth and vegetation growth of *Pinus radiata*. *For. Ecol. Manag.* 52, 179–199.
- Darrall, N.M., 1989. The effect of air pollutants on physiological processes in plants. *Plant Cell Environ.* 12, 1–30.

- De Kok, L.J., Tausz, M., 2001. The role of glutathione in plant reaction and adaptation to air pollutants. In: Grill, D., Tausz, M., De Kok, L.J. (Eds.), Significance of Glutathione to Plant Adaptation to the Environment. Kluwer Publishers, Amsterdam, pp. 185–208.
- Fenn, M.E., Poth, M.A., Aber, J.D., Baron, J.S., Bormann, B.T., Johnson, D.W., Lemly, A.D., McNulty, S.G., Ryan, D.F., Stottlemyer, R., 1996. Nitrogen excess in North American ecosystems: predisposing factors, ecosystem responses, and management strategies. *Ecol. Appl.* 8, 706–733.
- Fenn, M.E., Poth, M.A., 1999. Temporal and spatial trends in streamwater nitrate concentrations in the San Bernardino Mountains, southern California. *J. Environ. Qual.* 28, 822–836.
- Foyer, C.H., Lelandais, M., Kunert, K.J., 1994. Photooxidative stress in plants. *Physiol. Plant.* 92, 696–717.
- Gower, S.T., Haynes, B.E., Fassnacht, K.S., Running, S.W., Hunt Jr., E.R., 1993. Influence of fertilization on the allometric relations for two pines in contrasting environments. *Can. J. For. Res.* 23, 1704–1711.
- Grimes, H.D., Perkins, K.K., Boss, W.F., 1983. Ozone degrades into hydroxyl radical under physiological conditions. *Plant Physiol.* 72, 1016–1020.
- Grulke, N.E., Miller, P.R., Wilborn, R.D., Hahn, S., 1989. Photosynthetic response of giant sequoia seedlings and rooted branchlets of mature foliage to ozone fumigation. In: Olson, R.K., Lefohn, A.S. (Eds.), Effects of Air Pollution on Western Forests. Air and Waste Management Association, Pittsburgh, PA, pp. 429–441.
- Grulke, N.E., Miller, P.R., Scioli, D., 1996. Response of giant sequoia canopy foliage to elevated concentrations of atmospheric ozone. *Tree Physiol.* 16, 575–581.
- Grulke, N.E., Lee, E.H., 1997. Assessing visible ozone-induced foliar injury in ponderosa pine. *Can. J. For. Res.* 27, 1658–1668.
- Grulke, N.E., Andersen, C.P., Fenn, M.E., Miller, P.R., 1998. Ozone exposure and nitrogen deposition lowers root biomass of ponderosa pine in the San Bernardino Mountains, California. *Environ. Pollut.* 103, 63–73.
- Grulke, N.E., 1999. Physiological responses of ponderosa pine to gradients of environmental stressors. In: Miller, P.R., McBride, J.R. (Eds.), Oxidant Air Pollution Impacts in the Montane Forests of Southern California: A Case Study of the San Bernardino Mountains. In: Ecological Studies, Vol. 134. Springer-Verlag, New York, pp. 126–163.
- Grulke, N.E., Balduman, L., 1999. Deciduous conifers: high N deposition and O<sub>3</sub> exposure effects on growth and biomass allocation in ponderosa pine. *Water Air Soil Pollut.* 116, 235–248.
- Grulke, N.E., Retzlaff, W.A., 2001. Changes in physiological attributes of ponderosa pine from seedling to mature tree. *Tree Physiol.* 21, 275–286.
- Grulke, N.E., Preisler, H.K., Fann, C.C., Retzlaff, W.A., 2002a. A statistical approach to estimate ozone uptake of ponderosa pine in a mediterranean climate. *Environ. Pollut.* 119 (2), 163–175.
- Grulke, N.E., Preisler, H.K., Rose, C., Kirsch, J., Balduman, L., 2002b. O<sub>3</sub> uptake and drought stress effects on C acquisition of ponderosa pine in natural stands. *New Phytol.* 154, 621–632.
- Grulke, N.E., Johnson, R., Esperanza, A., Jones, D., Nguyen, T., Posch, S., Tausz, M., Canopy transpiration of Jeffrey pine in mesic and xeric microsites: O<sub>3</sub> uptake and injury response. *Trees*, in press.
- Heath, R.L., Taylor Jr., G.E., 1997. Physiological processes and plant responses to ozone exposure. In: Sandermann, H., Wellburn, A.R., Heath, R.L. (Eds.), Forest Decline and Ozone: A Comparison of Controlled Chamber and Field Experiments. In: Ecological Studies, Vol. 127. Springer-Verlag, New York, pp. 317–368.
- Hubbert, K., Beyers, J.L., Graham, R.C., 2001. Roles of weathered bedrock and soil in seasonal water relations of *Pinus jeffreyi* and *Arctostaphylos patula*. *Can. J. For. Res.* 31, 1947–1957.
- Kerstiens, G., Lenzian, K., 1989. Interactions between ozone and plant cuticles. I. Ozone deposition and permeability. *New Phytol.* 112, 13–19.

- Kiefer, J.W., Fenn, M.E., 1997. Using vector analysis to assess nitrogen status of ponderosa and Jeffrey pine along deposition gradients in forests of southern California. *For. Ecol. Manag.* 94, 47–55.
- Lee, E.H., Hogsett, W.E., 2001. Interpolation of temperature and non-urban ozone exposure at high spatial resolution over the western United States. *Climate Res.* 18, 163–179.
- Matyssek, R., Reich, P., Oren, R., Winner, R.E., 1995. Response mechanisms of conifers to air pollutants. In: Smith, W.K., Hinckley, T.M. (Eds.), *Ecophysiology of Coniferous Forests*. Academic Press, San Diego, CA, pp. 255–308.
- McDICK, J., Jarvis, P.G., Leahey, R.R.B., 1990. Influence of male cones on early season vegetative growth of *Pinus contorta* trees. *Tree Physiol.* 6, 105–117.
- Mehlhorn, H., Seufert, G., Schmidt, A., Kunert, K.J., 1986. Effects of SO<sub>2</sub> and O<sub>3</sub> on production of antioxidants in conifers. *Plant Physiol.* 82, 336–338.
- Mehlhorn, H., Tabner, B.J., Wellburn, A.R., 1990. Electron spin resonance: evidence for the formation of free radicals in plants exposed to ozone. *Physiol. Plant.* 79, 377–383.
- Miller, P.R., Parmeter, J.R., Taylor, O.C., Cardiff, E.A., 1963. Ozone injury to foliage of *Pinus ponderosa*. *Phytopathology* 53, 1072–1076.
- Miller, P.R., Longbotham, G.J., Longbotham, C.R., 1983. Sensitivity of selected western conifers to ozone. *Plant Disease* 67, 1113–1115.
- Miller, P.R., Stolte, K.W., Duriscoe, D.M., Pronos, J., 1996a. Evaluating ozone air pollution effects on pines in the western United States. Albany, California, Pacific Southwest Research Station General Technical Report, 155.
- Miller, P.R., Chow, J., Watson, J.G., 1996b. Assessment of acidic deposition and ozone effects on conifer forests in the San Bernardino Mountains. Final Report, Contract No. A032-180, California Air Resources Board, Research Division, 2020 L Street, Sacramento, CA 95814.
- Muir, P.S., Armentano, T.V., 1988. Evaluating oxidant injury to foliage of *Pinus ponderosa*: a comparison of methods. *Can. J. For. Res.* 18, 498–505.
- National Park Service, Air Resources Division, Sequoia National Park–Lower Kaweah data base, <http://www2.nature.nps.gov/ard/>.
- Patterson, M.T., Rundel, P.W., 1995. Stand characteristics of ozone-stressed populations of *Pinus jeffreyi* (*Pinaceae*): extent, development, and physiological consequences of visible injury. *Amer. J. Bot.* 82, 150–158.
- Pell, E.J., Dann, M.S., 1991. Multiple stress-induced foliar senescence and implications for whole-plant longevity. In: Mooney, H.A., Winner, W.E., Pell, E.J. (Eds.), *Responses of Plants to Multiple Stresses*. Academic Press, San Diego, CA, pp. 389–403.
- Pell, E.J., Sinn, J.P., Vinten Johansen, C., 1995. Nitrogen supply as a limiting factor determining the sensitivity of *Populus tremuloides* Michx. to ozone stress. *New Phytol.* 130, 437–446.
- Pell, E.J., Schlagnhauser, C.D., Artega, R.N., 1997. Ozone-induced oxidative stress: mechanisms of action and reaction. *Physiol. Plant.* 100, 264–273.
- Polle, A., 1998. Photochemical oxidants: uptake and detoxification mechanisms. In: De Kok, L.J., Stulen, I. (Eds.), *Responses of Plant Metabolism to Air Pollution and Global Change*. Backhuys Publishers, Leiden, The Netherlands, pp. 95–116.
- Pronos, J., Vogler, D.R., Smith, R.S., 1978. An evaluation of ozone injury to pines in the southern Sierra Nevada. Report 78-1. Pacific Southwest Region, USDA Forest Service, Forest Pest Management.
- Prus-Glowacki, W., Wojnicka-Poltorak, A., Oleksyn, J., Reich, P.B., 1999. Industrial pollutants tend to increase genetic diversity: evidence from field-grown European Scots pine populations. *Water Air Soil Pollut.* 116, 395–402.
- Reich, P.B., 1987. Quantifying plant response to ozone: A unifying theory. *Tree Physiol.* 3, 63–91.

- Retzlaff, W.A., Arthur, M.A., Grulke, N.E., Weinstein, D.A., Gollands, B., 2000. Use of a single tree simulation model to predict effects of ozone and drought on growth of a white fir tree. *Tree Physiol.* 20, 195–202.
- Salardino, D.H., Carroll, J.J., 1998. Correlation between ozone exposure and visible foliar injury in ponderosa and Jeffrey pines. *Atmos. Environ.* 32, 3001–3010.
- San Bernardino County Water District, Online data base: <http://www.co.san-bernardino.ca.us/t>.
- Schraudner, M., Langebartels, C., Sandermann, H., 1997. Changes in the biochemical status of plant cells induced by the environmental pollutant ozone. *Physiol. Plant.* 100, 274–280.
- Schweizer, B., Arndt, U., 1990. CO<sub>2</sub>/H<sub>2</sub>O gas exchange parameters of one- and two-year-old needles of spruce and fir. *Environ. Pollut.* 68, 275–292.
- Staszak, J., Grulke, N.E., Prus-Glowacki, W., Genetic structure of *Pinus ponderosa* (Dougl. ex Laws.) trees tolerant and sensitive to ozone. Unpublished data.
- Tausz, M., 2001. The role of glutathione in plant reaction and adaptation to natural stresses. In: Grill, D., Tausz, M., De Kok, L.J. (Eds.), *Significance of Glutathione to Plant Adaptation to the Environment*. Kluwer Publishers, Amsterdam, pp. 101–122.
- Tausz, M., Bytnerowicz, A., Arbaugh, M.J., Wonisch, A., Grill, D., 2001. Multivariate patterns of biochemical responses of *Pinus ponderosa* trees at field plots in the San Bernardino Mountains, southern California. *Tree Physiol.* 21, 329–336.
- Tausz, M., Herbinger, K., Posch, S., Grulke, N.E., 2002. Antioxidant status of *Pinus jeffreyi* needles from mesic and xeric microsites in early and late summer. *Phyton (Austria)* 42 (3), 201–207.
- Taylor, O.C., Kickert, R.N., McBride, J.R., Miller, P.R., Ohmart, C.P., Arkley, R.J., Dahlsten, D.L., Cobb Jr., R.W., Parmeter Jr., J.R., Luck, R.F., 1980. Photochemical oxidant air pollution effects on a mixed conifer forest ecosystem. Final Report, US Environmental Protection Agency, Office of Research and Development, 200 SW 35th Street, Corvallis, OR 97333.
- Temple, P.J., Riechers, G.H., 1995. Nitrogen allocation in ponderosa pine seedlings exposed to interacting ozone and drought stresses. *New Phytol.* 130, 97–104.
- Weber, J.A., Clark, C.S., Hogsett, W.E., 1993. Analysis of the relationships among O<sub>3</sub> uptake, conductance, and photosynthesis in needles of *Pinus ponderosa*. *Tree Physiol.* 13, 157–172.